

Prevalence of *Helicobacter Pylori* Infection among HIV-1 Infected Patients using Stool Antigen Tests in Jos, North-Central, Nigeria

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ABSTRACT

Background:

Helicobacter pylori (*H. pylori*) infection is common among humans and plays a major role in the etiology of peptic ulcer disease with significant morbidity in patients with HIV-1 on antiretroviral therapy. There are conflicting prevalence patterns of *H. pylori* in HIV-1 infected patients using various methods of detection. The noninvasive technique used for detection of *H. pylori* infection is inexpensive and convenient with no complications.

Materials and Methods:

We aimed to determine the prevalence of *H. pylori* infection among patients infected with HIV-1 on antiretroviral therapy using *H. pylori* stool antigen. 139 patients infected with HIV-1 were recruited, stool samples were collected and the *H. pylori* stool antigen (HpSA) test was used to detect *H. pylori* antigen.

Results:

46.8% of the respondents were positive for *H. pylori* and 53.2% were negative, 18 (13%) were men and 47 (33.8%) were women. HpSA is a relatively simple, inexpensive, and time-saving non-invasive test for the detection of *H. pylori* infections in patients infected with HIV-1.

Conclusion:

We also observed that the prevalence of *H. pylori* was low in these patients compared with the general population. However, more studies using *H. pylori* stool antigen test are needed in these patients in the North-Central, Nigeria to further evaluate the infection rate.

Keywords: Prevalence, *Helicobacter pylori*, Stool antigen, Enzyme immunoassay, HIV-1 infected

please cite this paper as:

Anejo-Okopi AJ, Audu O, Adaiche AR, Okojokwu OJ, Ali M, Adekwu A, Lar P, Smith SI. Prevalence of *Helicobacter Pylori* Infection among HIV-1 Infected patients using Stool Antigen Tests in Jos, North-Central, Nigeria. *Govaresh* 2016;21:55-63.

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Received: 29 Dec. 2015

Edited: 25 Feb. 2016

Accepted: 26 Feb. 2016

INTRODUCTION

Helicobacter pylori (*H. pylori*) infection is amongst the most common infections in humans strongly associated with peptic ulcer diseases, gastric cancer, and lymphomas(1). Approximately two-thirds of the world's population is infected with *H. pylori*; being more prevalent among older people in the United State of America, African Americans, Hispanics, and lower socio-economic groups(2,3). The pathogen was

first isolated and cultured from the antrum of patients with gastritis, and is a spiral-shaped bacterium commonly found in the gastric mucous layer or adherent to the epithelial lining of the stomach. The prevalence of *H. pylori* infection is more than 50% in developing countries, although the infection rates in most developed nations are dropping. The percent of *H. pylori*-associated gastric cancer is about 6.2% of all cancers, including 89% of non-cardia gastric cancer cases worldwide. However, the rates could be underestimated due to misdiagnosis or low sensitivity of detection of anti-*H. pylori* antibodies(4,5). The gastrointestinal tract(GIT) is the largest immunological site of the body where Human Immunodeficiency Virus type 1(HIV-1) is known to multiply greatly, and also serves as sanctuary site with attendant gut malfunctions(6). The common ulcer symptom is heart burning or gnawing in the epigastrium. The associated pain occurs typically when the stomach is left empty of food or drink, it can be between meals and in the early hours of the day. This pain may last from minutes to hours, may be relieved by eating or by taking antacids. The relationship between the bacteria and clinical manifestations is well known and it is associated with the presence of the *cagA* gene that triggers the pathogenicity island; such as Bab A, OIpA, and the carriers of the *vacA* gene. Studies have reported that only the *cagA* and *vacA* gene carriers are most likely to cause ulcer disease(7, 8). Smith et al has reported the relationship of *vacA* s1, *iceA1* and *cagA* as common genotypes of *H. pylori* infection and duodenal ulcers in Nigeria(9). Other studies have reported the prevalence of *H. pylori* in general population, children and also antibiotic susceptibility patterns in Nigeria(10), and developed countries(11,12).

It is well-known that *H. pylori* infection is mostly acquired during the early years of life and persists for several years in both developed and developing countries(13). About 25 million Americans suffer from peptic ulcer disease at some point in their lifetime, with little documented data in sub-Saharan Africa. More than 650,000 to 850,000 new cases of peptic ulcer disease and more than 1.5 million ulcer-related hospitalizations each year have been reported(14,15).

There are two broadly categorized available methods to detect *H. pylori* infection: invasive and

noninvasive methods. The invasive tests include: histology and culture. The cost and discomfort associated to the patients are very high and biopsy samples may be subject to errors related to sampling and interference of contaminated bacteria. The noninvasive tests include: urea breath test (UBT) and serology(16). The use of serological tests in most diagnostic laboratories and hospitals have some drawbacks; they do not discriminate between current and past infections, have low specificity and paucity of data on the specificity and sensitivity in HIV-infected individuals with immunodeficiency with perhaps altered production of antibody(17). The long-term retention of the antigen may cause false positive after the eradication of *H. pylori* because the *Helicobacter pylori* Stool Antigen (HPSA) tests does not distinguish between live and dead bacteria, however, comparative diagnostic methods of *H. pylori* have also reported the usefulness of HPSA in epidemiological findings in Nigeria(18).

The use of stool antigen as a noninvasive rapid test practical tool for detection of *H. pylori* infection is even more desirable in children. Stool antigen tests have recently been welcomed with great expectations as they are convenient and require little technical expertise to perform even in less equipped laboratories(19). However, the accuracy of stool antigen tests in different clinical set up outside of controlled studies posed a challenge, but HpSA GeneFront's ELISA VUETM (GeneFrontInc, USA) happens to be one of the most widely studied and it has shown to have acceptable performance in the detection of *H. pylori* infection(20). The stool polyclonal and monoclonal antigen tests have high sensitivity, specificity and accuracy in children and HIV-1 infected patients(21).

HIV-1 infected patients experience many forms of opportunistic infections including gastrointestinal symptoms(22). The exact role of *H. pylori* infection among HIV-1infected patients in gastro duodenal lesions might be different from the general population, and it remains unclear if upper gastrointestinal symptoms such as dyspepsia are highly active antiretroviral therapy (HAART) related adverse effects or as a result of *H. pylori* infection. The involvement of gastrointestinal tract may be because it represents the largest reservoir of HIV in the body with attendant

morphological changes in the upper-gastrointestinal tract mucosa and medication complications(23). Studies have shown that the prevalence of *H. pylori* infection in HIV-1 positive patients is remarkably low when compared with the general population(24). Review studies have shown low prevalence of *H. pylori* infection in HIV-1 infected patients therefore, the use of the stool antigen test is recommended for targeted high risk populations(25). We aimed to determine the prevalence of *H. pylori* using stool antigen by enzyme immunoassay among HIV-infected adults receiving highly active antiretroviral therapy (HAART) in Jos University Teaching Hospital (JUTH), Nigeria.

MATERIALS AND METHODS

Study Area and Population

The study was carried out at the HIV/AIDS treatment Centre of JUTH in collaboration with the AIDS Prevention Initiative in Nigeria (APIN) program. This clinic provides comprehensive HIV care services for the city of Jos and its metropolis. The clinic also serves as a referral center for health facilities in other Local Government Areas (LGAs) of the state, north central and neighboring states. The subjects used for this study include 139 HIV infected men and women aged ≥ 17 years receiving highly active antiretroviral therapy (HAART) at the centre.

Study design and study population

This was a cross-sectional study, and 139 HIV-1 infected volunteers accessing care at the JUTH antiretroviral clinic were enrolled. All patients included in the study provided written informed consent for the use of their data for research as approved by the institutional review board of Jos University Teaching Hospital (Health Research Ethic Committee) and were aged ≥ 17 years.

Data collection

Data collection lasted for four weeks. Consecutive eligible respondents seen over the study period were solicited to participate. Structured questionnaires was used to obtain information on socio-demographic characteristics of the respondents which included: age, sex, marital status, pet keeping, source of drinking water, sanitary practices, alcohol consumption, smoking habit, occupation, housing accommodation,

and signs of dyspepsia. Stool samples were collected from the patients with the aid of sterile stool containers with a spatula in them. The sterile spatula was attached to the cover and it was used to take the stool sample from tissue paper to avoid contamination from the toilet and it was covered immediately to avoid drying. Containers were labeled with the patients' identity number corresponding with the patient's identity on the questionnaire. Some samples were collected on same day while others were collected on the following day for convenient reasons. All samples were gathered and stored together at -80°C until enough samples were collected which lasted for a period of four weeks.

Laboratory Procedure: *Helicobacter pylori* Stool Antigen (HpSA) Test

An approximate peanut size of fresh stool sample were collected and stored at -20°C for analysis. The *H. pylori* polyclonal stool antigen-based GeneFront's ELISA VUETM (GeneFrontInc, Kukatpally, Hyderabad 500072, India) was performed according to manufacturer's instructions. The test is quantitative, and based on a sandwiched enzyme immunoassay for antigen detection with purified *H. pylori* antibody coated on the surface of micro-wells. One hundred and ninety two coated strips were placed into the holder and $10\mu\text{l}$ of treated sample calibrators and controls were dispensed into the appropriate wells. Air bubble was removed by tapping the holder from the liquid and it was mixed well. The stripe and the holder were incubated at room temperature for 30 mins. First, $100\mu\text{l}$ of a diluted stool sample ($10\mu\text{l}$ stool in 0.5 ml sample diluent) and thereafter, peroxidase-conjugated polyclonal antibody solution were added to the wells and incubated for 30 mins at room temperature. The unbound material was removed by washing, and washing was repeated 3 times with washing buffer. $100\mu\text{l}$ of enzyme conjugate was dispensed into each well and incubated at room temperature for 30 mins. After addition of a TMB chromogenic substrate, 100 ml solution was dispensed into each well and was incubated for 15 mins at room temperature and $100\mu\text{l}$ of stop solution was added to stop reaction, and the intensity of the color generated is proportional to the amount of antigen in the sample. The optical density was read at 450nm by a spectrophotometer compared in a parallel manner with calibrator and controls. The

cutoff OD value for sero-positivity was >20ng/ml and negative <15ng/ml.

Statistical Analysis

Statistical analysis was done using Statistical Package for Social Sciences (SPSS) version 16.0. Standard descriptive statistics were used to examine the sero-prevalence, while associations between socio-demographic characteristics, health status, and variables of potential exposure (environmental and lifestyle conditions), and *H. pylori* sero-positivity of HIV-1 infected patients were examined using bivariate analysis and Chi-square. $P \leq 0.05$ was considered as statistically significant.

RESULT

Of 139 diagnosed HIV-1 infected patients who participated in the study, the prevalence rate of *H. pylori* infection was 46.8%, 18 (13.0%) were men and 38 (33.8%) were women.

With respect to age category, 17-26 year-olds had the highest frequency (15.1%, n=21), followed by 27-36 year-olds (12.2%, n=17), 37-46 year-olds (11.0%, n=15), and ≥ 41 year-olds (8.6%, n=12). 38 (27.3%) were married predominated and 27 (19.4%) were single. Most (25.2%, n=35) were unemployed, 21 (15.1%) worked in the private sector or were traders, and 9 (6.5%) were public servants. 27 (19.4%) patients had a history of alcoholic consumption and 7 (5.0%) smoke cigarettes. 29 (21.0%), 11 (8.0%), and 25 (18.0%) of the patients used the pit latrine, water system, and bush for sanitary purposes, respectively ($P=0.05$). 45 (32.4%) had room accommodation and 20 (14.4%) had flat apartments. 14 (10.1%), 12 (8.6%), and 39 (28.0%) patients used tap water, borehole, and well for drinking water, respectively. Amongst those with signs of dyspepsia, abdominal pain was seen in 35 (25.2%) patients, followed by belching in 11 (8.0%), vomiting 2 (1.4%), heart burn 10 (7.2%), and bloating in 2 (1.4%) patients. 5 (3.6%) patients had no symptoms. 28.8% of the patients kept had a pet ($p=0.30$). Although this association was not statistically significant, but the odds of having *H. pylori* infection was about one and half times more in those keeping pets compared with those not keeping pets (OR=95%, CI=1.4, 0.73-2.82). There were no significant differences between *H. pylori* infection

in the patients' with respect to age, marital status, occupation, alcohol consumption, smoking habit, housing accommodation, and drinking water source, (table 1).

DISCUSSION

The International Agency for Research on Cancer (IARC) reported that *H. pylori* is a type I carcinogen, or a cancer definite in humans. This report was based almost exclusively on epidemiological evidence, though controversial. It has been argued in some quarters that *H. pylori* is only a risk factor(26). Gastric cancers are a leading cause of cancer morbidity and mortality worldwide; with adenocarcinomas arising from gastric glands accounting for 90% of incident cases(27). The stomach wall is being protected from the gastric juice by thick mucus layer of the stomach lining. *H. pylori* take advantage of this layer by living in the mucus lining. Earlier reported findings showed that more than half of the world's population is infected with *H. pylori*, which is acquired during the early years of life. Actual infection rates vary from nation to nation; the developing countries have much higher infection rates (90%) than the developed countries (1.2-12%)(28). *H.pylori* infection has been known to be associated with gastritis, duodenal ulcer, gastric cancer, and mucosa associated lymphoid tissue lymphoma(17,34). *H.pylori* infection may be asymptomatic; therefore, we included in our study patients with and without digestive complaints, since it may provide better chances for the pathogen detection. Our results showed low prevalence of *H. pylori* infection (46.8%) in HIV-1 patients which differs remarkably from that previously reported study for HIV-negative adults in Nigeria (70.3%) (18) and Brazil (82.0%) (24). This study was stool antigen based, but we observed low prevalence against the earlier reported study in Nigeria 36.7%(29). Similarly, studies have reported different prevalent rates using sera (*H.pylori* IgG antibodies) in Ibadan (82.7-94.5%), Abeokuta (47.8%), Ile-Ife (73.0 %) and Lagos (93.6%), all in South western Nigeria(30-32). Holcombe et al who used histology, Haematoxillin and Eosin, with modified Giemsa staining of antral biopsies in Maiduguri found a prevalence rate (84%) of *H. pylori* among dyspeptic patients(33).

Also earlier reported findings showed that the

Table 1: Socio-demographic characteristics of Helicobacter pylori infection among HIV-1 Patients attending Jos University Teaching Hospital, Nigeria (n=139).

Variables	Number positive (%)	Number negative (%)	P value
Age Category			0.570
17-26	21(15.1)	18(13.0)	
27-36	17(12.2)	14(10.1)	
37-46	15(11.0)	13(9.3)	
≥41	12(8.6)	29(21.0)	
Sex			0.600
Male	18(13.0)	21(15.1)	
Female	47(33.8)	53(38.1)	
Marital status			0.060
Single	27(19.4)	35(25.2)	
Married	38(27.3)	39(28.0)	
Occupation			0.600
Public servant	9(6.5)	38(27.3)	
Private/Trading	21(15.1)	27(19.4)	
Unemployed	35(25.2)	9(6.5)	
Alcohol consumption			0.100
Yes	27(19.4)	30(21.6)	
No	38(27.3)	44(31.6)	
Smoking habit			0.200
Never	58(42.0)	66(47.5)	
Currently	7(5.0)	7(5.0)	
Past	0(0.0)	1(0.7)	
Sanitary practices			0.050
Pit	29(21.0)	26(18.7)	
Water system	11(8.0)	40(28.8)	
Bush	25(18.0)	8(5.7)	
Housing accommodation			0.090
Room	45(32.4)	40(28.8)	
Flat	20(14.4)	34(24.5)	
Source of drinking water			0.060
Tap	14(10.1)	34(24.5)	
Borehole	12(8.6)	19(13.7)	
Well	39(28.0)	21(15.1)	
Signs of dyspepsia			0.070
Abdominal pain	35(25.2)	38(27.3)	
Belching	11(8.0)	15(11.0)	
Vomiting	2(1.4)	8(5.7)	
Heartburn	10(7.2)	4(3.0)	
Bloating	2(1.4)	7(5.0)	
None	5(3.6)	2(1.4)	
Pet keeping			0.30
Yes	40(28.8)	39(28.0)	
No	25(18.0)	35(25.2)	

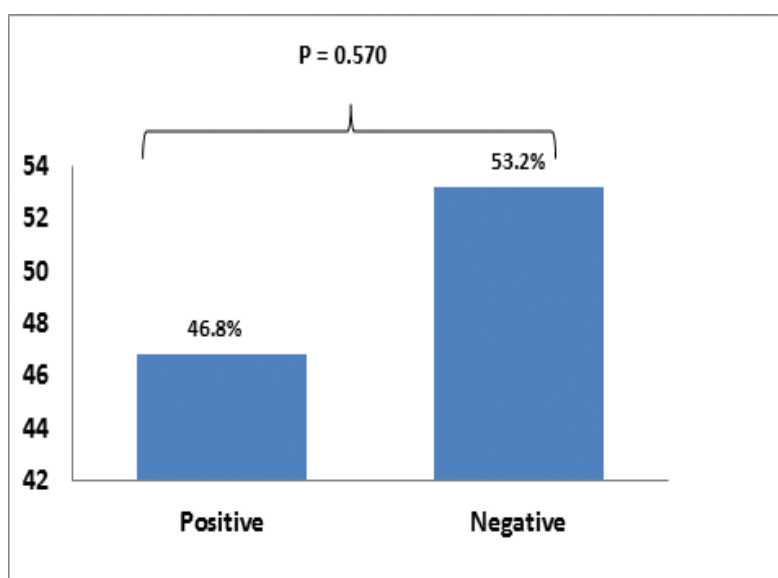


Fig. 1: Prevalence of Helicobacter pylori infection among HIV-1 Patients attending Jos University Teaching Hospital, Nigeria

prevalence of *H. pylori* infection has been decreasing over the years in the general population(34) but this assertion cannot be easily verified because of the variability of research methodologies employed. However, studies from Kenya, Cameroon and Nigeria also reported higher prevalence of 60% to 71% in dyspepsia patients and in apparently healthy individuals. Similar higher prevalence was reported in western Nigerian in patients with duodenal ulcers (>90%)(35-38).

Our study is consistent with similar reported findings in HIV-1 infected patients from Western Nigeria (47.4%)(39), though lower prevalence (36.7%) of *H. pylori* infection using HpSA had also been reported in Nigeria among HIV-1 negative individuals(29). Studies from other parts of the world have reported lower prevalence in HIV infected individuals (11.36%, and 22.1%)(40,41).

This study revealed that high proportion (15.1%) of participants aged 17-26 years were infected with *H. pylori* as compared to older age categories, and this was consistent with earlier reported findings in Nigeria(36), and Turkey(42). Similarly, the study observed high proportions of *H. pylori* infection among the women with no significant risk of association, though larger percentage of women had participated compared to men.

Poor sanitary practices and signs of dyspepsia among the participants were found to have a significant association with *H. pylori*. The high prevalence of *H. pylori* infection among those that used pit latrine and bush could be attributed to poor socioeconomic status. However, studies have documented a relation between low socio-economic status and the high rates of *H. pylori* infection in other parts of the world and Nigeria(34,37). The significant association of signs of dyspepsia among HIV-1 infected patients we observed in our study was consistent with earlier reported studies in Nigerian adults on suppressive antiretroviral therapy(39). The role of *H. pylori* in gastrointestinal disturbances and dyspepsia related symptoms has been documented which is as a result of immunosuppression and therapy indications(6,41).

In our study, the lack of a significant association may be due to a small sample size. This was similar to earlier findings that acquisition of *H. pylori* infection is common through poor environmental sanitation practices. In particular, animals and contaminated or untreated water have been implicated as potential sources of *H. pylori* infection. The possibility that *H. pylori* may be a zoonosis first arose following reported findings that the prevalence of *H. pylori* infection in abattoir and meat workers had significantly increased as compared to other people not involved in handling

animals or animal products(43). Dore et al have also reported a positive association between the prevalence of *H. pylori* in Sardinian shepherds and contact with sheep and sheep dogs(44). In their study, 98% of shepherds were shown to be infected with *H. pylori*, a prevalence that was significantly higher than those of other family members who did not have regular contact with sheep (73%). Moreover, the isolation of *H. pylori* from the stomach of an entire colony of pathogen-free animals suggested that animals represent an important reservoir of *H. pylori*(45). Similarly, unhygienic sources of water are important routes of *H. pylori* transmission; our study observed that consumption of well water or tap water had a near significant association to *H. pylori* infection which was consistent to earlier findings in healthy Nigerian children with poor socio-economic status(46) and other developing countries(28).

The use of ¹³C urea breath test and other invasive methods are cumbersome, expensive, relatively unavailable tests in many resource limited countries where infrastructure posed a challenge(47), but noninvasive methods such as stool antigen test has proven to be effective in children and immunocompromised individuals indicating low prevalence compared to serological tests with higher outcomes. However, *H. pylori* infection is mostly acquired during childhood which persists in adulthood, but it is also suggestive that the HIV-infected patients studied might have been exposed to the pathogen early in life and this infection could be lost on account of several healthy life style and ART treatment after acquired HIV-1. Similarly, the *H. pylori* gastric load might be decreased in the HIV-1 patients due to interplay of CD4 cell count, gastric mucosal colonization by other pathogens and indiscriminate use of either antibiotics or proton-pump inhibitors (PPIs) which is known to modify the mucosal environment. The use of antibiotics for treatment or prophylaxis on initiation of HIV-1 infected patients and possible cofounders could suggest low prevalence and or misdiagnosis of *H. pylori*(48).

CONCLUSION

The prevalence of *H. pylori* infection in HIV-1 infected patients on suppressive therapy was low using HpSA ELISA stool antigen test which is

effective in detecting true presence of the antigen and acute infection. However, an interesting finding on the association of marital status, housing accommodation, source of drinking water and signs of dyspepsia were observed. Other factors such as; sanitary practices and pet keeping were associated with *H. pylori* infection though not statistically significant, but suggests involvement of poor socio-economic status of *H. pylori* co-infected with HIV-1 infected individuals. Though the association of *H. pylori* through poor sanitary practices and pet keeping has been documented in this study, therefore attention should be given to fecal-oral transmission of *H. pylori* infection as well as other possible routes of transmission in order to reduce infection among low social class of people. There is need for more studies of large sample sizes to be carried out using stool antigen test because they are simple, non-invasive, relatively inexpensive and reliable assays in the diagnosis of *H. pylori* infection.

ACKNOWLEDGMENT

This publication was facilitated, in part, by the US Department of Health and Human Services, Health Resources and services Administration (U51HA02522- 01-01) which Funded HIV/AIDS treatment and care services at JUTH, APIN, Jos. We sincerely appreciate the JUTH APIN management for support and permission to use the patients for the study.

CONFLICT OF INTERESTS

Authors have declared that no conflicting interests exist.

REFERENCES

1. Marshall B, Adams CP. Helicobacter pylori: A Nobel pursuit?. *Can J Gastroenterol* 2008; 22:895-6.
2. Huang JQ, Zheng GF, Sumanac K, Irvine EJ, Hunt RH. Meta-analysis of the relationship between cagA seropositivity and gastric cancer. *Gastroenterology* 2003;125:1636-44.
3. Dore PM, Malaty MH, Graham YD, Fanciulli G, Delitala G, Realdi G. Risk Factors associated with Helicobacter pylori infection among children in a defined geographic area. *Clin Infect Dis* 2002;35: 240-5.

4. Testerman LT, Morris J. Beyond the stomach: An updated view of *Helicobacter pylori* pathogenesis, diagnosis, and treatment. *World J Gastroenterol* 2014;20:12781-808.
5. Plummer M, Franceschi S, Vignat J, Forman D, de Martel C. Global burden of gastric cancer attributable to *Helicobacter pylori*. *Int J Cancer* 2015;136:487-90.
6. Chehter ZE, Catapani RW, Margeotto BF, Germini D, Henriques CA, Waisberg J. *Helicobacter pylori* in the Era of Highly Active Antiretroviral Therapy (HAART): A Review. *JSM Gastroenterol Hepatol* 2014;2:1026-30.
7. Bashinskaya B, Nahed BV, Redjal N, Kahle KT, Walcott BP. Trends in peptic ulcer disease and identification of *Helicobacter pylori* as a causative organism: population-based estimates from the US nationwide inpatient sample. *J Glob Infect Dis* 2011;3:366-70.
8. Smith IS, Kirsch C, Oyedeji SK, Arigbabu OA, Cokerk OA, Bayerdoffer E, et.al. Prevalence of *Helicobacter pylori* vacA, cagA and iceA genotypes in Nigerian patients with duodenal ulcer disease. *J Med Microbiol* 2002;51:851-4.
9. Holcombe C, Tsimiri S, Eldridge J, Jones MD. Prevalence of antibody to *Helicobacter pylori* in children in northern Nigeria. *Trans R Soc Trop Med Hyg* 1993;87:19-21.
10. Ani AE, Malu OA, Onah AJ, Queiroz MD, Kirschner G, Rocha AG. Antimicrobial susceptibility test of *Helicobacter pylori* isolated from Jos, Nigeria. *Trans R Soc Trop Med Hyg* 1999;93:659-61.
11. Osato SM, Reddy R, Graham YD. Metronidazole and clarithromycin resistance amongst *Helicobacter pylori* isolates from a large metropolitan hospital in the United States. *Int J Antimicrob Agents* 1999; 12:341-7.
12. Mendonca S, Ecclissato C, Sartori MS, Godoy AP, Guerzoni AR, Degger MJ, et.al. Prevalence of *Helicobacter pylori* resistance to metronidazole, clarithromycin, amoxicillin, tetracycline, and furazolidone in Brazil. *Helicobacter* 2000;5:79-83.
13. Torres J, Pérez-Pérez G, Goodman KJ, Atherton JC, Gold BD, Harris PR, et.al. A comprehensive review of the natural history of *Helicobacter pylori* infection in children. *Arch Med Res* 2000;31:431-69.
14. Perry S, Sanchez ML, Yang S, Haggerty DT, Hurst P, Perez-Perez G, et. al. Gastroenteritis and Transmission of *Helicobacter pylori* Infection in Households. *Emerg Infect Dis* 2006;12:1701-8.
15. Volk J, Parsonnet J. Epidemiology of Gastric Cancer and *Helicobacter pylori*, The Biology of Gastric Cancers Springer Science New York, 2009; pp25-57.
16. Makristathis A, Pasching E, Schtze K, Wimmer M, Rotter ML, Hirschl AM. Detection of *Helicobacter pylori* in stool specimens by PCR and antigen enzyme immunoassay. *J Clin Microbiol* 1998;36:2772-4.
17. Lepper PM, Moricke A, Vogt K, Bode G, Trautmann M. Comparison of different criteria for interpretation of immunoglobulin G immunoblotting results for diagnosis of *Helicobacter pylori* infection. *Clin Diagn Lab Immunol* 2004;11:569-76.
18. Smith SI, Oyedeji KS, Goodluck HA, Fowora MA, Anomneze E, Lesi OA. The Use of *Helicobacter Pylori* Stool Antigen test for the Diagnosis of *Helicobacter Pylori* in Lagos, Nigeria. *West Indian Med J* 2011; 60:33-6.
19. Mansour-Ghanaei F, Mashhour MY, Joukar F, Sedigh M, Bagher-Zadeh AH, Jafarshad R. Prevalence of *Helicobacter Pylori* Infection among Children in Rasht, Northern Iran. *Middle East J Dig Dis* 2009;1:84-8.
20. Gisbert JP, Pajares JM. Stool antigen test for the diagnosis of *Helicobacter pylori* infection: a systematic review. *Helicobacter* 2004;9: 347-68.
21. Kato S, Ozawa K, Okuda M, Nakayama Y, Yoshimura N, Konno M, et. al. Multicenter comparison of rapid lateral flow stool antigen immunoassay and stool antigen enzyme immunoassay for the diagnosis of *Helicobacter pylori* infection in children. *Helicobacter* 2004;9:669-73.
22. Skwara P, Mach T, Tomaszewska R. Morphological changes of gastric mucosa in HIV-infected patients. *HIV and AIDS Rev* 2002; 2:47-51.
23. Werneck-Silva AL, Prado IB. Dyspepsia in HIV-infected patients' under highly active antiretroviral therapy. *J Gastroenterol Hepatol* 2007;22:1712-6.
24. Fialho BCA, Braga-Neto BM, Guerra JCE, Fialho MNA, Fernandes CK, Sun LMJ, et al. Low prevalence of *H. pylori* Infection in HIV Positive Patients in the Northeast of Brazil. *BMC Gastroenterol* 2011;11:1-5.
25. Panos GZ, Xirouchakis E, Tzias V, Charatsis G, Bliziotis IA, Doulgeroglou V, et.al. *Helicobacter pylori* infection in symptomatic HIVseropositive and seronegative patients: a case-control study. *AIDS Res*

- Hum Retroviruses* 2007;23:709-12.
26. Suerbaum S, Michetti P. Helicobacter pylori infection. *N Engl J Med* 2002;347:1175-86.
 27. Coleman MP, Esteve J, Damiecki P, Arslan A, Renard H. Trends in Cancer incidence and mortality. *IARC Sci Publ* 1993;121:1-806.
 28. Frenck WR, Clemens J. Helicobacter in the developing world. *Microbes Infect* 2003;5:705-13.
 29. Smith IS, Omonigbehin AE, Goodluck AH, Abdulkareem BF, Onyekwere AC, Rieder G, et al. Diagnostic methods for the detection of Helicobacter pylori in Nigeria. *Trop Gastroenterol* 2010;31:113-5.
 30. Adeniyi BA, Otegbayo JA, Lawal TO, Oluwasola AO, Odaibo GN, Okolo C, et al. Prevalence of Helicobacter pylori infection among dyspepsia patients in Ibadan, South West Nigeria. *Afr J Microbiol Res* 2012;6:3399-402.
 31. Oluwasola AO, Ola SO, Saliu L, Solanke TF. Helicobacter pylori infection in South Nigerians: a serological study of dyspeptic patients and healthy individuals. *West Afr J Med* 2002;21:138-41.
 32. Abdollahi A, Shoar S, Jafari S, Emadi-Kochak H. Seroprevalence of helicobacter pylori in human immunodeficiency virus-positive Patients and its correlation with CD4+ Lymphocyte Count. *Niger Med J* 2014;55:67-72.
 33. Holcombe M, Umar H, Lucas SB, Kaluba J. Low incidence of clinically significant gastroduodenal pathology despite a high incidence of H. pylori infection. *Trans Roy Soc Trop Med Hyg* 1994;88: 569-571.
 34. Özden A, Bozdayı G, Özkan M. Changes in the sero-epidemiological pattern of Helicobacter pylori infection over the last 10 years in Turkey. *Turk J Gastroenterol* 2001;15:156-8.
 35. Shmuelly H, Obure S, Passaro JD, Abuksis G, Yahav J, Fraser G, et.al. Dyspepsia Symptoms and Helicobacter pylori Infection, Nakuru, Kenya. *Emerg Infect Dis* 2003;9:103-7.
 36. Otegbay A, Oluwasola OA, Yakubu A, Odiabo GN, Olaleye OD. Helicobacter pylori serology and evaluation of gastroduodenal disease in Nigerians with dyspepsia. *Afr J Clin Exper Microbiol* 2001; 5:131-8.
 37. Mynepalli CKS, Maureen O, Mumuni A. Prevalence of Helicobacter pylori and hygiene practices among public secondary school students in Ikeja Local Government Area, Lagos, Nigeria. *J Health* 2014;6:250-8.
 38. Smith SI, Oyedeji KS, Arigbabu A, Anomneze EE, Chibututu CC, Atimomo CA. Prevalence of H. pylori in patients with gastritis and peptic ulcer in Western, Nigerian. *Br J Biomed Sci* 2001;58:97-100.
 39. Ejilude O, Akinduti PA, Idowu M, Ogunbileje JO, Akinbo JA. HIV and Helicobacter Pylori Co-Infection In Dyspeptic Patients In Abeokuta, Nigeria. *New York Sci J* 2011;4:1-5.
 40. Şincu NI, Chiriac CL, Fodor AM. Helicobacter pylori infection in HIV-positive patients with digestive complaints. *Revista Romana de Medicina de Laborator* 2014;22;1-11.
 41. Sud A, Ray P, Bhasin D, Wanchu A, Bambery P, Singh S. Helicobacter pylori in Indian HIV infected patients. *Trop Gastroenterol* 2002;23:79-81.
 42. Pounder RE, Ng D. The prevalence of Helicobacter pylori infection in different countries. *Aliment Pharmacol Ther* 1995;9: 33-9.
 43. Vaira D, D'Anastasio C, Holton J, Dowsett JF, Londei M, Bertoni F. et. al. Campylobacter pylori in abattoir workers. *Lancet* 1998;725:726.
 44. Dore MP, Sepulveda AR, Osato MS, Realdi G, Graham, D.Y. Helicobacter pylori in sheep. *Lancet* 1999;354:132.
 45. Handt LK, Fox JG, Yan LL, Shen Z, Pouch WJ, Ngai D, et.al. Diagnosis of Helicobacter pylori infection in a colony of rhesus monkeys (Macaca mulatta). *J Clin Microbiol* 1997;165:168.
 46. Senbanjo OI, Oshikoya AK, Njokanma FO. Helicobacter pylori associated with breastfeeding, nutritional status and recurrent abdominal pain in healthy Nigerian children. *J Infect Dev Ctries* 2014; 8:448-53.
 47. Lu CY, Kuo FC, Wang SW. The clinical applications and accuracy of two rapid near-patient tests in detecting Helicobacter pylori infection. *Diagn Microbiol Infect Dis* 2006;56:241-6.
 48. Lichterfeld M, Lorenz C, Nischalke HD, Scheurlen C, Sauerbruch T, Rockstroh JK. Decreased prevalence of Helicobacter pylori infection in HIV patients with AIDS defining diseases. *Z Gastroenterol* 2002;40:11-4.